

Crystal Structure of Adenylylsulfate Reductase from *Desulfovibrio gigas* Suggests a Potential Self-regulation Mechanism Involving the C-terminus of the β -subunit

Yuan-Lan Chiang (江元郎)^{1,2}, Yin-Cheng Hsieh (謝殷程)^{1,2},
Jou-Yin Fan (方柔伊)^{1,2}, En-Hong Liu (劉恩鴻)^{1,2}, Phimonphan Chuankhayan (邱康妍)¹,
Jeyakanthan Jeyaraman (桀肯特)¹, Min-Yih Liu (劉明毅)¹, Sunney I. Chan (陳長謙)⁴,
and Chun-Jung Chen (陳俊榮)^{1,3,5}

¹National Synchrotron Radiation Research Center, Hsinchu, Taiwan

²Institute of Bioinformatics and Structural Biology,
National Tsing Hua University, Hsinchu, Taiwan

³Department of Physics, National Tsing Hua University, Hsinchu, Taiwan

⁴Institute of Chemistry, Academia Sinica, Taipei, Taiwan

⁵Institute of Biotechnology, National Cheng Kung University, Tainan, Taiwan

Adenylylsulfate reductase (adenosine 5'-phosphosulfate reductase, APS reductase or APSR) plays a key role in catalyzing APS to sulfite in dissimilatory sulfate reduction. Here, we report the crystal structure of APSR from *Desulfovibrio gigas* at 3.1 Å resolution. Different from the $\alpha_2\beta_2$ heterotetramer of the *A. fulgidus*, the overall structure of APSR from *D. gigas* comprises six $\alpha\beta$ -heterodimers that form a hexameric structure. The flavin adenine dinucleotide (FAD) is non-covalently attached to the α -subunit, and two [4Fe-4S] clusters are enveloped by cluster-binding motifs. The substrate-binding channel in *D. gigas* is wider compared to *A. fulgidus* because of shifts in the loop (326-332) and the α -helix (289-299) in the α -subunit. The positively charged residue Arg160 in the structure of *D. gigas* likely replaces the role of Arg83 of *A. fulgidus* for the recognition of substrates. The C-terminal segment of the β -subunit wraps around the α -subunit to form a functional unit, with the C-terminal loop inserted into the active-site channel of the α -subunit from another $\alpha\beta$ -heterodimer. Electrostatic interactions between the substrate-binding residue Arg282 in the α -subunit and Asp159 on the C-terminus of the β -subunit affect the binding of the substrate. Alignment of the APSR sequences from *D. gigas* and *A. fulgidus* shows the largest differences toward the C-terminal of the β -subunit, and structural comparison reveals the notable differences at the C-terminal, activity site and other regions. The disulfide Cys156–Cys162 stabilizes the C-terminal loop of the β -subunit and is crucial for oligomerization. Dynamic light scattering and ultracentrifugation measurements reveal multiple forms of the APSR upon the addition of adenosine monophosphate (AMP), indicating that AMP binding dissociates the inactive hexamer into functional dimers presumably by switching C-terminus of β -subunit away from the active site. The crystal structure of APSR, together with its oligomerization properties, suggests that APSR from sulfate-reducing bacteria might self-regulate its activity through the C-terminus of the β -subunit [1].

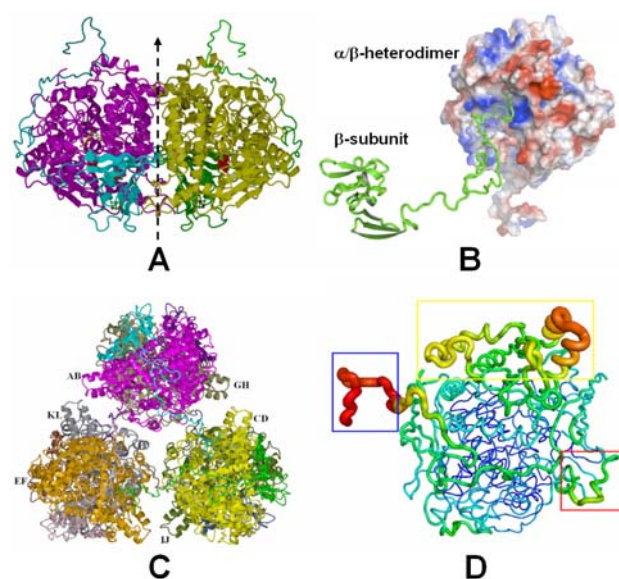


Fig. 1: Structure of APS reductase hexamer from *D. gigas*. (A) The structure of $\alpha_2\beta_2$ heterotetramer from *D. gigas*. The $\alpha\beta$ -heterodimer performs a rotation about a 2-fold axis with another $\alpha\beta$ -heterodimer to form a tightly contacted $\alpha_2\beta_2$ heterotetramer. (B) Interactions between the $\alpha\beta$ -heterodimers and C-terminus of another β -subunit. The long C-terminal loop of the β -subunit (shown in ribbon; green) plugs into the active channel of another $\alpha\beta$ -heterodimer (shown in electrostatic surface). (C) A view of the hexamer structure. Three $\alpha_2\beta_2$ heterotetramers contact each other through the C-termini of the β -subunits to form a hexamer containing six $\alpha\beta$ -heterodimers. (D) The B-factor labeled structure of $\alpha\beta$ -heterodimer. The structure exhibits a high B-factor with red color and the larger caliber. The blue box shows the high B-factor at the C-terminus of the β -subunit. The electron-accepting site on the β -subunit that is suggested to interact with an unknown electron donor shows a higher B-factor in the red box. The capping domain in the α -subunit also shows a higher B-factor in the yellow box.

Reference

- [1] Y.-L. Chiang, Y.-C. Hsieh, J.-Y. Fang, E.-H. Liu, Y.-C. Huang, P. Chuankhayan, J. Jeyaraman, M.-Y. Liu, S. I. Chan, and C.-J. Chen, *J. Bacteriol.* **191**, 7597 (2009).